2021 MOB X-Ray Boot Camp

Fall 2021 | 3 Week Course

Crystallization and Diffraction

Syllabus and Calendar

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CYRSTALLIZATION AND DIFFRACTION

Practical course in Protein Crystallization and X-Ray Diffraction
Session: Lecture: 40 min; Lab: 120 min; Q&A: 20 min; 3 weeks 5 sessions

GOALS

The practical course in crystallization and x-ray diffraction is a precursor to a graduate level macromolecular crystallography special class taught every other year. In this course, we will cover concepts of crystals, symmetry, crystallization, x-ray generation and safety, x-ray diffraction, cryo safety and technique, data collection and processing, Linux o/s, computer graphics, and data visualization. In this practical course, more emphasis will be given to hands-on training and lab work than the full-fledged graduate level course.

At the end of this course, you will learn basic concepts in:

- Symmetry, space groups, lattice, and unit cell
- Need for crystals, cryogenic temperatures, and X-Rays
- Manual and robotic crystallization set-up and
- Crystal harvesting, imaging, and loading
- Cryogenic safety and cryo-crystallography
- X-Ray generation and safety
- X-Ray diffraction and data collection
- Linux operating system and running programs
- Computer graphics and data analysis

CALENDER

MOB 2020 Cohort Boot Camp Schedule

August 2021 | XRF Schedule in purple font

<table>
<thead>
<tr>
<th>Monday</th>
<th>Tuesday</th>
<th>Wednesday</th>
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<td>02-Aug</td>
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<td>1-5:Solution Prep</td>
<td>8-12:Data Collection</td>
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<td>16-Aug</td>
<td>17-Aug</td>
<td>18-Aug</td>
<td>19-Aug</td>
<td>20-Aug</td>
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<td>1-5:Cryo Prep</td>
<td>1-5: Data Process</td>
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8-12 Data Analysis | Classes Start

XRF Sessions are in purple font (8-12: AM session and 1-5: PM session)
COURSE

Session: We will meet for three hours per session for five sessions. The five sessions will be spread over three weeks. The session will consist of 20 minutes lecture followed by 60 minutes of lab-work or hands on training followed by 10 minutes of Question and Answer session.

Course: The total course length is for three weeks with two sessions per week. We expect the course will have 4-5 students per session and all lab supplies will be provided to the students.

SYLLABUS

1. Symmetry, lattices, crystals, and crystallization
2. Crystals and protein crystallization set-up
3. Crystallization observation and optimization
4. X-Rays, radiation safety and X-Ray generation
5. Cryogenics safety and cryocrystallography
6. X-Ray Diffraction set-up and data collection
7. Linux operating system and computer graphics
8. Data processing and visualization and wrap-up

Sessions highlighted in blue will involve more hands-on training time and sessions highlighted in green will involve more lecture time than indicated in the original description.

RESOURCES

Requirements

- Wet lab bench work experience
- Computer skills using Windows/Mac/Linux operating systems
- Basic knowledge about macromolecules and their properties

Resources

- Kevin Cowtan’s Crystallography Teaching Materials
- Cambridge University Crystallography Course
- A folder containing all the required materials will be given to the students on the first day

CONTACT INFORMATION

- Dr. Thayumana Somasundaram
- Phone: 1-850-644-6448
- E-mail: tsomasundaram@fsu.edu
- URL: biophysics.fsu.edu/soma/projects/core-facilities-workshop/
DETAILED SYLLABUS

1. Symmetry, lattices, crystals, and crystallization
In this session, we will cover basic ideas of symmetry: point groups, space groups, symmetry elements, Bravais lattices, and chiral molecules and their symmetry. Then we will study about crystals of macromolecular materials. We will compare and contrast crystal growth techniques. The lab portion of the session will involve learning about hanging-drop and sitting-drop vapor diffusion experiments. We will also cover Commercial Crystallization Screens. [2021-Mod-0_Introduction.pptx, 2021-Mod-1_Symm+Charzn.pptx, 2021-Mod-2_Crystln.pptx]

2. Crystals and protein crystallization set-up
In this session, we will start to learn about macromolecular crystals and their characteristics. We will learn about de novo crystallization. We will also learn about commercial screens available for proteins, nucleic acids from various vendors. We will also learn about crystal screening services available for testing multiple conditions. The lab portion of the session will involve preparing protein solutions (lysozyme, buffers, and precipitating agents. We will be setting up manual hanging-drop and robotic sitting-drop vapor diffusion crystallization trays. [2021-Mod-2_Crystln.pptx, 2021-Mod-1+2_Crystallization.docx]

3. Crystallization observation and optimization
In this session, we will optically observe and score the crystal trays we have set-up during the last session. Based on the original scores modified crystallization conditions will be prepared and additional trays will be set-up for future observation and harvesting. We will robotically image crystals, manipulate and mount crystals. Previously prepared crystals will be mounted into glass capillaries, and cryo loops. [2021-Mod-1+2_Crystallization.docx]

4. X-Rays, radiation safety, and X-Ray generation
In this session, we will cover topics pertaining to need for X-Ray radiation, why we need to carry out X-Ray diffraction to obtain atomic resolution images of macromolecules. We will cover basics of X-Ray safety. How X-Rays are produced both at home and at a synchrotron station. The lab portion of the session will involve tour the home X-Ray Diffraction laboratory and exploring various components involved in collecting the data. [2021-Mod-3_X-Ray+Cryo_Safety.pptx]

5. Cryogenic safety and cryo-crystallography
In this session, we will start with safety dealing with liquid nitrogen. We will then move on to the need for low-temperature (cryo-crystallography) crystallography. The advantages and disadvantages of low-temp work will be discussed. During the lab portion of the session, we will learn how to mount a crystal in a cryo loop. We will also learn how to flash-cool the crystal in a loop under bulk liquid nitrogen bowl and low-temp flowing nitrogen gas. [2021-Mod-3_X-Ray+Cryo_Safety.pptx, 2021-Mod-3+4_CryoCrystallography.docx]

6. X-Ray diffraction set-up and data collection
In this session crystals in glass capillaries cryo loops will be mounted on to the X-Ray diffraction machine. Crystal alignment and detector specific adjustments will be taught. In the lab portion of the session data from ambient or low-temperature crystal, diffraction data will be collected.

[2021_XRF_SOP.pdf; XRayView demonstration]

7. Linux operating system and computer graphics

In this session, basic Linux operating system commands and procedures will be introduced. Introduction to computer graphics and macromolecular graphics programs will be introduced. [Chimera, PDB]

8. Data processing and visualization

Using a shared account data collected during the previous session will be visualized, processed, and simple interpretation of the data will be carried out. [Phenix: Xtriage, Molecular Replacement, Refinement].

MANUALS AND HANDOUTS

0. Introduction, syllabus, and schedule

Presentation [pptx]

1. Symmetry, space groups, lattices, and unit cell

Presentation [pptx]

2. Crystals and protein crystallization set-up

Presentation [pptx] | Crystallization Module [pdf]

3. Crystallization observation and optimization

Presentation [pptx] | Crystallization Module [pdf]

4. X-Rays, radiation safety, and X-Ray generation

Presentation [pptx]

5. Cryogenic safety and cryo-crystallography

Presentation [pptx] | Cryo Crystallography [pdf]

6. X-Ray diffraction set-up and data collection

Presentation [pdf] | Data Collection Module [pdf]

7. Linux operating system and data processing

Presentation | Data Processing Module [pdf]

8. Protein Data Bank Data processing and data visualization


Updated: July 06, 2021.